SHORT COMMUNICATION

STUDIES IN THE CRUCIFERAE: STEROLS IN POLLEN OF BRASSICA NAPUS L.

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(Received 30 March 1968)

Abstract—Using GLC with mass spectrometry the main sterols of pollen from *Brassica napus L. f. annua* (cv. Giant English rape) have been shown to be 24-methylenecholesterol and 24-ethylidenecholesterol.

INTRODUCTION

STUDIES carried out at Gif-Sur-Yvette have shown that pollens may contain unusual sterols, and sterols not found in other parts of the plant. Thus it has been shown that 24-methylene-cholesterol is often the principal sterol of pollen, occurring in Zea mays L., Pyrus malus L., Salix spp. and Cistus ladanifera L. Cholesterol is the principal sterol in pollen of Hypochoeris radicata L.^{2,3} A new sterol, pollinastanol, was isolated from pollen of Castanea vulgaris Lam. and Corylus avellana L.⁴ Corylus avellana pollen also contained mono- and diunsaturated C₂₇, C₂₈ and C₂₉ sterols. During work involving studies of the effects of Plasmodiophora brassicae woron in the Cruciferae, it was found that marked changes in sterol composition occurred in plants at different stages of development. As a part of a study of this effect, pollen of Brassica napus L. f. annua (cv. Giant English rape) was collected and the sterol composition studied.

RESULTS AND DISCUSSION

Sterols were isolated in the usual way 5 after the pollen grains had been ruptured as far as possible by grinding in light petroleum suspension. Figure 1 shows the gas chromatograms obtained from this sterol fraction as trimethylsilyl ethers using SE-30 and OV-17 stationary phases. The two main peaks (3 and 6) correspond in mobility with the derivatives of 24-methylenecholesterol and Δ^5 -avenasterol (24-ethylidenecholesterol) respectively. The identity of these two compounds was confirmed by combined gas chromatography-mass spectrometry (CG-MS) using SE-30 stationary phase. The obtained mass spectra of the trimethylsilyl ethers were identical with previously described spectra for these two compounds. The spectrum from peak 5+6 showed two extra ions indicating the presence of some β -sitosterol, and peak 5 in the OV-17 trace corresponds in retention time to β -sitosterol.

¹ M. BARBIER, M. F. HUGEL and E. LEDERER, Bull. Soc. Chim. Biol. 42, 91 (1960).

² M. Devys and M. Barbier, Compt. Rend. Acad. Sci., (Paris) 261, 4901 (1965).

³ M. Devys and M. BARBIER, Phytochem. 5, 1031 (1966).

⁴ M. F. Hugel, Ann. Abeille 8, 309 (1965); Chem. Abs. 64, 19718 (1966).

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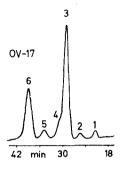
⁶ B. A. KNIGHTS, J. Gas Chromatog. 5, 273 (1967).

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GLC and mass spectral data for peak 1 indicated that this compound was cholesterol and the mass spectrum of peak 2 was consistent with the possibility of this compound being pollinastanol. The identity of compound 4 is uncertain, but mass spectral data suggest it corresponds to a C_{29} diunsaturated sterol with a 4-methyl group.

The relative percentages of each sterol, obtained by triangulation of peaks in the GLC traces, are respectively:

Compound 1, (cholesterol) 2.8 per cent; compound 2, (pollinastanol?) 2.4 per cent; compound 3, (24-methylenecholesterol) 56 per cent; compound 4(?) 3 per cent; compound 5, (β -sitosterol) 4.2 per cent; compound 6, (24-ethylidenecholesterol $\equiv \Delta^5$ -avenasterol) 31.6 per cent.



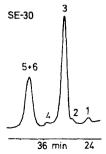


Fig. 1. Pollen sterol trimethylsilyl ethers.

Operating conditions 9 ft columns; 3 per cent OV-17 256°, N₂ carrier gas—68 ml/min; 1 per cent SE-30 250°, N₂ carrier gas—60 ml/min.

EXPERIMENTAL

Pollen from flowering plants was collected by detaching the flowers and scraping the anthers with a knife blade. The yield of sterol, isolated in the usual way,⁵ was approximately 0·5 per cent. GLC was carried out in a Pye 104 model 14 chromatograph using 9 ft columns packed with either 1 per cent SE-30 coated on Gas Chrom P⁷ or 3 per cent OV-17 coated on Gas Chrom Q (Applied Science Laboratories pretested packing). The GC-MS analysis was obtained using an LKB-9000 gas chromatograph-mass spectrometer equipped with a 10 ft 1 per cent SE-30 column.

Acknowledgements—This work was supported by an A.R.C. research grant. An LKB-9000 GC-mass spectrometer was purchased under grant number B/SR/2398. The provision of this facility by Drs. C, J. W. Brooks and G. Eglinton and the technical assistance of Miss H. Humphrys and Miss J. Malcolm is gratefully acknowledged.

⁷ E. C. HORNING, W. J. A. VANDENHEUVEL and B. G. CREECH, in *Methods of Biochemical Analysis* (edited by D. GLICK), Vol. XI, p. 69, Wiley, New York (1963).